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Nucleosides, Nucleotides and Nucleic Acids

Publication details, including instructions for authors and subscription information:

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Oxidative Cyclization of 2',3'-O-Isopropylidene-Adenosines into 5'-O,8-Cycloadenosines with Lead Tetraacetate: Remarkable Effect of N⁶-Substituents on the Oxidation

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To cite this Article Kitade, Yukio , Makino, Tohru , Hirota, Kosaku and Maki, Yoshifumi(1992) 'Oxidative Cyclization of 2',3'-O-Isopropylidene-Adenosines into 5'-O,8-Cycloadenosines with Lead Tetraacetate: Remarkable Effect of N⁶-Substituents on the Oxidation', *Nucleosides, Nucleotides and Nucleic Acids*, 11: 2, 365 — 372

To link to this Article: DOI: 10.1080/07328319208021710

URL: <http://dx.doi.org/10.1080/07328319208021710>

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**OXIDATIVE CYCLIZATION OF 2',3'-O-ISOPROPYLIDENE-
ADENOSINES INTO 5'-O,8-CYCLOADENOSINES WITH LEAD
TETRAACETATE: REMARKABLE EFFECT OF *N*⁶-SUBSTITUENTS
ON THE OXIDATION†**

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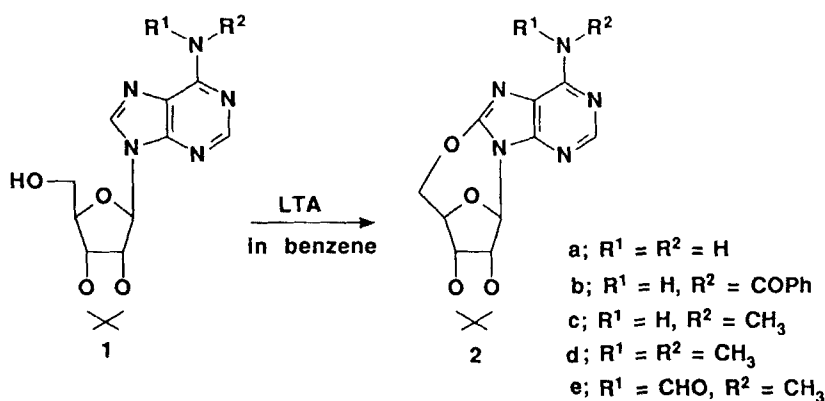
Abstract

Oxidation of 2',3'-*O*-isopropylideneadenosines (**1**) with lead tetraacetate (LTA) in dry benzene resulted in the formation of the corresponding 5'-*O*,8-cyclo-2',3'-*O*-isopropylideneadenosines (**2**), which has a new methodological implication for the chemical modification of adenosines. The occurrence of the oxidative cyclization was remarkably affected by the nature of *N*⁶-substituents: *N*⁶-benzoyl substitution prominently accelerated the oxidative cyclization in comparison with none and dimethyl substitutions. In the oxidation of *N*⁶,*N*⁶-dimethyladenosine (**1d**), an intriguing oxidative demethylation was observed.

Introduction

Since cyclonucleosides are generally regarded as versatile intermediates for the preparation of biologically important nucleosides and as tool for conformational studies of nucleosides,^{1,2} many efforts have been made to explore a new methodology for the preparation of cyclonucleosides. Research in this field still has many untouched possibilities to explore. For example, although 5'-*O*,8-cyclo-2',3'-*O*-isopropylideneadenosine (**2a**) was synthesized by a conventional method involving the intramolecular cyclization of the corresponding 8-bromoadenosine in the presence of base,³ we have found that 2',3'-*O*-isopropylideneadenosine (**1a**) undergoes an unprecedented-

†This contribution is dedicated to the late Professor Tohru Ueda with our deepest sympathy.



Scheme 1

ed oxidative cyclization to give **2a** upon treatment with oxidants such as lead tetraacetate (LTA),⁴ cupric chloride,⁵ and *N*-halogenosuccinimide,⁶ and by irradiation in the presence of pyrimido[5,4-*g*]pteridinetetrone 5-oxide.⁷ The reaction mode of these intramolecular cyclizations obviously contains either a one-electron oxidation in two stages or a two-electron oxidation, and provides a clue for the development of new methods for the chemical modification of adenosines.

In this paper, we wish to describe in full detail⁸ the oxidative cyclization of 2',3'-*O*-isopropylideneadenosines into the corresponding 5'-*O*,8-cycloadenosines with LTA, in particular, the remarkable substituent effect of *N*⁶-substituents on the reaction, which sheds some light on the oxidation mode.

Results and Discussion

A stoichiometric study showed that the use of 1.2 equivalent of LTA is required for the smooth conversion of the 2',3'-*O*-isopropylidene derivative (**1a**) into the cycloadenosine (**2a**). Thus, refluxing **1a** with LTA (1.2 eq.) in dry benzene under argon for 2.5 h afforded **2a** in 93 % yield. The cycloadenosine (**2a**) was identical in all respects with an authentic sample.³ The oxidative cyclization of adenosine itself with LTA did not give the expected 5'-*O*,8-cycloadenosine under the conditions employed. Therefore, 2',3'-*O*-isopropylidene protection of adenosines is a prerequisite for the oxidative cyclization.

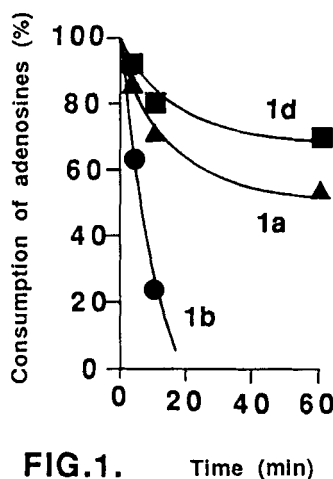
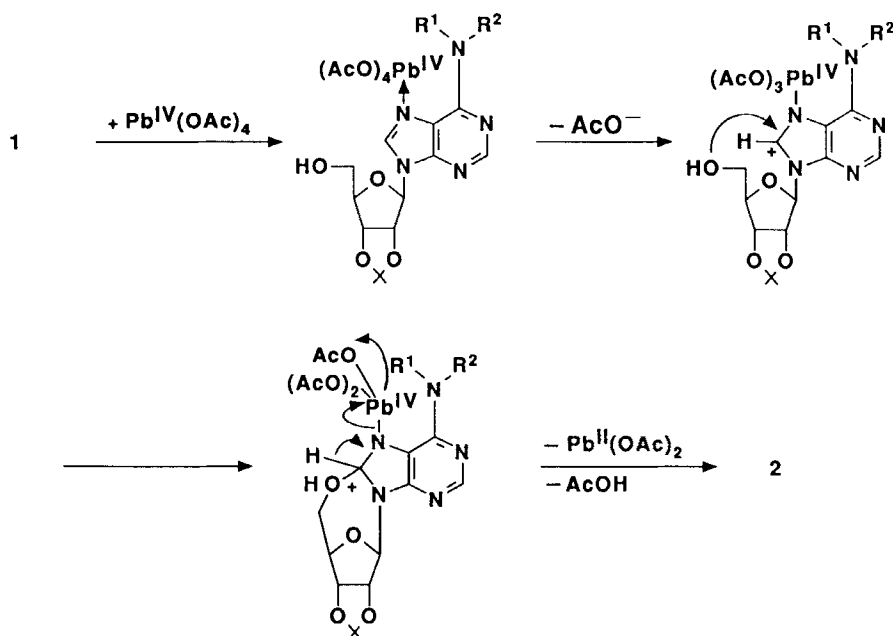


FIG.1. Time (min)

When N^6 -benzoyladenosine derivative (**1b**) was employed as a substrate, the oxidative cyclization smoothly proceeded to give the corresponding 5'-O,8-cycloadenosine (**2b**), quantitatively. Under the analogous conditions, the oxidative cyclization of N^6 -methyladenosine (**1c**) and N^6,N^6 -dimethyladenosine (**1d**) gave the corresponding 5'-O,8-cycloadenosines (**2c**) and (**2d**) in 65% and 40% yield, respectively. In the case of **1d**, the formation of small amounts of other oxidized products was observed (*vide infra*). The structures of **2b-d** were fully supported by mass, $^1\text{H-NMR}$, and microanalytical results.

Figure 1 shows consumption of **1a**, **1b**, and **1d** in the reaction with LTA as a function of reaction time, indicating clearly that the ease of the oxidation is in the order of **1b** > **1a** > **1d**. The prominent substituent effect of the N^6 -benzoyl group on the present conversion is unequivocal. The relative increase of the nucleophilicity of the imidazole ring nitrogen (N^7) in comparison with the pyrimidine nitrogen (N^1) by virtue of the introduction of N^6 -acyl group in the adenosines has been manifested by concrete observations, e.g. contrary to the adenosines, protonation and alkylation of N^6 -acyladenosines predominantly occur at the N^7 -position rather than the N^1 -position.⁹ Thus, the accelerative effect of the N^6 -benzoyl group on the present conversion can be explained in terms of the preferential coordination of lead ion at the N^7 -position of **1d** for the oxidative cyclization. A conceivable reaction sequence for the present oxidative cyclization by lead ion is outlined in the manner of an intramolecular oxidative nucleophilic substitution¹¹ involving a two-electron oxidation as depicted in Scheme 2.



Scheme 2

Reaction of the N^6,N^6 -dimethyladenosine (**1d**) with LTA (1.2 eq.) under reflux in dry benzene for 2 h resulted in the formation of the corresponding cycloadenosine (**2d**)(40%) and demethylated products (**2c**)(5%) and (**1c**)(2%) together with a trace amount of N^6 -formyl- N^6 -methylcycloadenosine (**2e**). The structure of **2e** was fully supported by its $^1\text{H-NMR}$ and high-resolution mass spectrum, e.g. the NMR spectrum of **2e** showed the presence of only one methyl group (δ 3.55, s, 3H) and the N -formyl group (δ 10.10, s, 1H). When a mixture of **1d** and LTA (3 eq.) in large excess in benzene was refluxed for 1.5 h, the yields of demethylated products increased to some extent: **2c** (26%) and **1c** (8%). In the oxidation of **1d** under an oxygen atmosphere, no distinct change in both the products distribution and the consumption rate of **1d** was observed. Analogous treatment of **2d** with LTA gave a mixture of **2c** and **2e** in 60% and ca. 3% yield, respectively.

Endo and Zemlicka¹² have reported that the oxidation of N^6,N^6 -dimethyl-2',3',5'-tri- O -acyladeniosine with ruthenium tetroxide gives selectively the corresponding N^6 -formyl- N^6 -methyl derivative. Oxidation of the N^6,N^6 -dimethyl

group in **1d** with LTA has an interesting mechanistic implication in connection with the previous observation.¹² Special attention will be paid to the occurrence of competitive oxidation with LTA in both the *N*⁶,*N*⁶-dimethyl group and the imidazole ring in **1d** in order to clarify the detailed oxidative mechanism.

In summary, the present study provided a basic observation which is applicable to the development of new methodology for the chemical modification of purine nucleosides.

Experimental Section

Melting points were determined with a Yanagimoto micro melting point apparatus and are uncorrected. Elemental analyses were carried out at the microanalytical laboratory of our university. All UV measurements were carried out with a Shimadzu 260 spectrometer. ¹H-NMR spectra were recorded at 270 MHz on a JEOL JNX-270 spectrometer in CDCl₃. Chemical shifts are quoted in parts per million (s = singlet, d = doublet, t = triplet, and m = multiplet). Mass spectra (MS) were measured at 70 eV with a JEOL JMS-D300 spectrometer. TLC for the assay of adenosines was performed on Silica gel 60 plates (Merk, art 5715) by using chloroform-methanol (5:1) as an eluent and TLC-scanning was carried out with a Shimadzu CS-9000 dual-wavelength flying-spot scanner (detection, the corresponding maximum peak of adenosines). Column chromatography was carried on silica gel (Wako gel C-200).

5'-O,8-Cyclo-2',3'-O-isopropylideneadenosine (2a). A mixture of 2',3'-O-isopropylideneadenosine (**1a**) (154 mg, 0.5 mmol) and lead tetraacetate (90%) (LTA) (296 mg, 0.6 mmol) in dry benzene (60 mL) was refluxed for 2.5 h under argon. The resulting precipitate was removed by filtration and the filtrate was evaporated under reduced pressure. The residue was chromatographed on a silica gel column with chloroform-methanol (30:1). The appropriate fractions containing the product were collected and the solvent was removed under reduced pressure to give **2a** (143 mg, 93%), which was identical in every respect with an authentic sample prepared by base-catalyzed cyclization of 8-bromo-2',3'-O-isopropylideneadenosine.³

***N*⁶-Benzoyl-5'-O,8-cyclo-2',3'-O-isopropylideneadenosine (2b).** A mixture of *N*⁶-benzoyl-2',3'-O-isopropylideneadenosine (**1b**) (123 mg, 0.3 mmol) and LTA (90%) (177 mg, 0.36 mmol) in dry benzene (10 mL) was

refluxed for 15 min. under argon. The resulting precipitate was removed by filtration and the filtrate was evaporated under reduced pressure. The residue was chromatographed on a silica gel column with chloroform-methanol (40:1). The appropriate fractions containing the product were collected and the solvent was removed under reduced pressure to give **2b** (120 mg); m.p. 234-236 °C. ¹H NMR: δ 1.36 (s, CH₃), 1.57 (s, CH₃), 4.26 (d, 1H, J = 13.2 Hz, C₅-H), 4.55 (d, 1H, J = 13.2 Hz, C₅-H), 4.76 (m, 2H, C₃-H and C₄-H), 5.09 (d, 1H, J = 5.86 Hz, C₂-H), 6.49 (s, 1H, C₁-H), 7.48 (t, 2H, J = 7.32 Hz, C₂-H), 7.57 (t, 1H, J = 7.32 Hz), and 8.01 (d, 2H, J = 7.32 Hz). Anal. Calcd. for C, 58.68; H, 4.68; N, 17.11: Found C, 58.45; H, 4.75; N, 16.91; High-resolution mass spectrum: 409.1417 (M⁺, C₂₀H₁₉N₅O₅ requires 409.1414).

5'-O,8-Cyclo-2',3'-O-isopropylidene-N⁶-methyladenosine (2c). A mixture of 2',3'-O-isopropylidene-N⁶-methyladenosine (**1c**) (438 mg, 1.36 mmol) and LTA (90%) (802 mg, 1.63 mmol) in dry benzene (15 mL) was refluxed for 2 h under argon. The resulting precipitate was removed by filtration and the filtrate was evaporated under reduced pressure. The residue was chromatographed on a silica gel column with chloroform-methanol (30:1). The appropriate fractions containing the product were collected and the solvent was removed under reduced pressure to give **2c** (282 mg, 65%). ¹H NMR: δ 1.35 (s, CH₃), 1.56 (s, CH₃), 3.22 (d, 3H, N⁶-H), 4.17 (d, 1H, J = 12.82 Hz, C₅-H), 4.47 (d, 1H, J = 12.82 Hz, C₅-H), 4.71 (br, 1H, C₄-H), 4.75 (d, 1H, J = 5.55 Hz, C₂-H), 5.08 (d, 1H, J = 5.55 Hz, C₂-H), 5.86 (s, 1H, N⁶-H), 6.40 (s, 1H, C₁-H), and 8.35 (s, 1H, C₂-H); High-resolution mass spectrum: 319.1285 (M⁺, C₁₄H₁₇N₅O₄ requires 319.1284).

Reaction of 2',3'-O-isopropylidene-N⁶,N⁶-dimethyladenosine (1d) with LTA. A mixture of **1d** (100 mg, 0.3 mmol) and LTA (90%) (177 mg, 0.36 mmol) in dry benzene (10 mL) was refluxed for 2 h under argon. The resulting precipitate was removed by filtration and the filtrate was evaporated under reduced pressure. The residue was chromatographed on a silica gel column with chloroform-methanol (40:1). The first fraction gave 5'-O,8-cyclo-2',3'-O-isopropylidene-N⁶-formyl-N⁶-methyladenosine (**2e**) (< 1%). ¹H NMR: δ 1.25 (s, CH₃), 1.37 (s, CH₃), 3.55 (s, 6H, N⁶-CH₃), 4.26 (d, 1H, J = 12.21 Hz, C₅-H), 4.45 (d, 1H, J = 12.21 Hz and 2.20 Hz, C₅-H), 4.77 (m, 2H, C₃-H and C₄-H), 6.53 (s, 1H, C₁-H), 8.63 (s, 1H, C₂-H), and 10.18 (1H, s, N⁶-CHO); High-resolution mass spectrum: 347.1216 (M⁺, C₁₅H₁₇N₅O₅ requires 347.1218).

The second fraction gave 5'-O,8-cyclo-2',3'-O-isopropylidene-*N*⁶,*N*⁶-dimethyladenosine (**2d**) (40 mg, 40%). ¹H NMR: δ 1.36 (s, CH₃), 1.56 (s, CH₃), 3.43 (s, 6H, *N*⁶-dimethyl), 4.16 (d, 1H, J = 13.19 Hz, C₅-H), 4.45 (d, 1H, J = 13.19 Hz, C₅-H), 4.75 (d, 1H, J = 5.37 Hz, C₃-H), 5.08 (d, 1H, J = 5.55 Hz, C₂-H), 6.42 (s, 1H, C₁-H), and 8.29 (s, 1H, C₂-H); High-resolution mass spectrum: 333.1452 (M⁺, C₁₅H₁₉N₅O₄ requires 333.1452). The third fraction gave **2c** (5%), which was identical with the sample prepared above. The fourth fraction gave **1c** (2%), which was identical with the authentic sample.

Reaction of 2d with LTA. A mixture of **2d** (100 mg, 0.3 mmol) and LTA (90%) (177 mg, 0.36 mmol) in dry benzene (15 mL) was refluxed for 1 h under argon. The resulting precipitate was removed by filtration and the filtrate was evaporated under reduced pressure. The residue was chromatographed on a silica gel column with chloroform-methanol (40:1). The early-eluting fraction gave **2c** (60 mg, 60%), which was identical in every respect with the sample obtained above. The later-eluting fraction gave **2e** (3 mg, 3%), which was identical in every respect with the sample obtained above.

Measurement of Consumption of 2',3'-O-isopropylideneadenosines (1a, 1b, and 1d) in the oxidation with LTA. A mixture of adenosines (**1a**, **1b**, or **1d**) (5mM) and LTA (6mM) in dry benzene (20 mL) was refluxed under argon. Consumption of the adenosines (**1a**, **1b**, and **1d**) was followed spectrophotometrically with a TLC scanner and was calculated by $A/A_0 \times 100$ (A_0 = the initial peak area of adenosines, A = the peak area of adenosines in the time course of the oxidation).

Acknowledgments

This work was partly supported by a Grant-in-Aid for Scientific Research on Priority Areas No. 03242104 from the Ministry of Education, Science and Culture, Japan.

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Received 8/22/91

Accepted 12/4/91